

Antimicrobial and Cytotoxic Activities of the Crude Extracts of *Polyalthia Simiarum*

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Abstract

The extractives of *Polyalthia simiarum* (Annonaceae) were subjected to antimicrobial screening and brine shrimp lethality bioassay. In case of antimicrobial screening petroleum ether and ethyl acetate extracts exhibited promising antibacterial activity, while the petroleum ether extract demonstrated highest cytotoxicity with LC₅₀ of 1.91 µg/ml in the brine shrimp lethality bioassay.

1. Introduction

Polyalthia simiarum Hook. f. Thom. locally known as Arjan, (Family- Annonaceae) is a very tall tree which grows in Cox's Bazar hillside of Bangladesh. It is also grown in India, Bhutan, Myanmar, Thailand, Laos and Vietnam.¹ The species of this genus are reported to have cytotoxic and antimicrobial,² anticancer³ and antimalarial⁴ activities. The species of this genus is used for the treatment of skin disease, fever, diabetes, hypertension and helminthiasis.⁵ As part of our studies of medicinal plants of Bangladesh, we herein, report the antimicrobial and cytotoxic activities of the crude extracts of *P. simiarum*.

II. Materials and Methods

The stem bark of *P. simiarum* was collected from Mirpur, Dhaka in the month of June 2008 and identified by Mr. Sarder Nasir Uddin, Scientific Officer, Bangladesh National Herbarium, Dhaka, where a voucher specimen (DACB-34201) representing this collection has been deposited. The

air dried powdered plant material (700 g) was sequentially extracted in a Soxhlet apparatus with petroleum ether (60-80°C) followed by ethyl acetate. The extractives were filtered through fresh cotton plug and followed by whatman no.1 filter paper. The filtrate were then concentrated by a Buchii rotavapor at low temperature and pressure and afforded pet-ether extract (PE, 3.5g), ethyl acetate extract (EA, 2.5g). The antimicrobial activities of the crude extracts were determined by the disc diffusion method,^{6,7,8} against the bacterial strains listed in Table-1. These were collected as pure cultures from the Institute of Nutrition and Food Science (INFS), University of Dhaka, Bangladesh. Here Kanamycin (30 µg/disc) was used as the standard. The pet-ether and ethyl acetate extracts were dissolved separately in chloroform and applied to sterile discs at a concentration of 400 µg/disc and carefully dried to evaporate the residual solvent.

Table. 1. Antimicrobial activity of *P.simiarum* extracts(400µg/disc) and kanamycin (30 µg/disc)

Test microorganisms	Diameter of zone of inhibition (mm)		KAN
	PE	EA	
Gram Positive			
<i>Bacillus cereus</i>	25	23	35
<i>B. megaterium</i>	28	27	38
<i>B. subtilis</i>	25	25	36
<i>Staphylococcus aureus</i>	24	23	35
<i>Sarcina lutea</i>	25	27	36
Gram Negative			
<i>Escherichia coli</i>	-	-	37
<i>Pseudomonas aeruginosa</i>	23	25	27
<i>Salmonella paratyphi</i>	23	25	36
<i>S. typhi</i>	20	21	35
<i>Shigella boydii</i>	24	25	36
<i>Vibrio mimicus</i>	21	23	37
<i>V. parahemolyticus</i>	21	21	35
Fungi			
<i>Candida albicans</i>	27	28	35
<i>Aspergillus niger</i>	25	25	35
<i>Sacharomyces cerevacaee</i>	25	28	38

PE: Pet ether extract, EA: Ethyl acetate extract; KAN: kanamycin. The zone of inhibition less than 8 mm was considered inactive.

Table 2. LC₅₀ data of *P. simiarum* extracts and vincristine sulfate.

Samples	LC ₅₀ µg/ml
VS.	0.32
PE	1.91
EA	3.65

The values of LC₅₀ were expressed in µg/ml. VS: Vincristine Sulfate (Std); PE: Pet-ether extract; EA: Ethyl acetate extract. For cytotoxicity screening, DMSO solutions of the petroleum ether and ethyl acetate extracts were applied against *Artemia salina*⁹ in one-day *in vivo* assay. For the experiment 4 mg of each of the pet-ether and ethyl acetate extracts were dissolved in DMSO and solutions of varying concentrations 400, 200, 100, 50, 25, 12.50, 6.25, 3.125, 1.563, 0.781(µg/ml) were made by serial dilution technique for each extract.

III. Results and Discussions

The crude pet-ether and ethyl acetate extracts showed promising antibacterial activity with the average zone of inhibition of 20-28mm and 21-28mm, respectively, at 400µg/disc. The pet-ether extract showed the highest activity against the growth of *B. megaterium* having the zone of inhibition of 28mm. Besides the growth of *B. cereus*(25mm), *B. subtilis*(25mm), *S. lutea*(25mm), *S. boydii*(24mm), *S. aureus*(24mm), *P. aeruginosa*(23mm), *S. paratyphi*(23mm), *S. dysenteriae*(23mm) exhibited prominent activity. In the case of fungi, the average zone of inhibition was found to be 25-28mm. At the same time, the ethyl acetate extracts also inhibited the growth of *B. megaterium*(27mm), *S. lutea*(27mm), *B. subtilis*(25mm), *p. aeruginosa*(25mm), *S. paratyphi*(25mm), *.boydii*(25mm), *S. dysenteriae*(25mm), *B. cereus*(23mm), *S. aureus*(23mm) and *V. mimicus*(23mm) significantly. The same extract also exhibited high inhibitory activity against the growth of fungal strains. Following the procedure of Meyer,⁹ the lethality of the pet-ether (PE) and ethyl acetate (EA) extracts to brine shrimp were evaluated on *A. salina* after 24 hours⁹ of exposure the sample and the positive control, Vincristine sulphate(VS). The LC₅₀ were found to be 0.32, 1.91, 3.65 µg/ml for VS, PE and EA extracts respectively. The cytotoxicity exhibited by the crude extracts were promising and this clearly indicates the presence of potent bioactive compounds.

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1. Khanam M, Rahman MM.(2002) Annonaceae, in: Khan MS, Rahman MM.(Eds), *Flora of Bangladesh*, **52**, Bangladesh National Herbarium, Ministry of Environment and Forest, Government of the People's Republic of Bangladesh 52.
2. Hasan CM, MA Hossain, MA Rashid, Connolly JD.(1994) Constituents from *Polyalthia longifolia* var *pendula*, *Fitoterapia*, **65**, 283-284.
3. Sashidhara KV, SP Sing, J. Sarkar, S. Sinha (2009) Cytotoxic clerodane diterpinoids from the leaves of *Polyalthia longifolia* *Natural Product Research: formerly Natural Product Letters*, 1-8.
4. Kanokmedhakul S, K. Kanokmedhakul, Lekpluom R.(2007) Bioactive constituents of the roots of *Polyalthia cerasoides*, *Journal of Natural Products*, **70**, 1536-1538.
5. Awadhes Kumar¹ and GS Solanki² (2008) The chemistry, pharmacologic and therapeutic applications of *Polyalthia longifolia* *Primate Conservation*, **23**, 97-105.
6. Baur, A.W., W.M.M., Kirby, J.C. Sherris and M. Turek, 1966. Antibiotic susceptibility testing by a standardized single disk method. *Am.J. Clin. Pathol.* **45**, 493-496.
7. Gazi, H.R., Kabir, S., Rahman, M. S., Chowdhury, A. M. S., Begum, B. and Rashid, M. A., 2007. Antimicrobial and cytotoxic activities of the crude extracts of *Hopea scaphula*, *Dhaka Univ. J. Pharm. Sci.* **6(2)**: 131-133.
8. Nahar, K., M.G.U Khan, M.S Rahman, B. Begum and M.A., Rashid, 2008. Antimicrobial and cytotoxic activities of *Bryophyllum daigremontianum*. *Dhaka Univ. J. Pharm. Sci.* **7(1)**: 99-101.
9. Meyer, B.N., N.R. Ferringni, J. E., Puam, L.B., Lacobsen, D.E. Nichols and J. L., McLaughlin, 1982. Brine shrimp a convenient general bioassay for active constituents. *Planta Med.* **45**, 31-32.

